

# SN CHICKEN RT-PCR DETECTION KIT

## Description of the Product:

The SN Chicken RT-PCR detection kit constitutes a ready to use system for multiple detection (presence or absence) using polymerase chain reactions (PCR) in the Real Time PCR machine. The Specific Master mix contains reagents and enzymes for the specific amplification of Chicken genes to generate amplified product and for the direct detection of the specific amplicon in fluorescence channel Cy5 of the Real Time PCR & the reference gene / Inhibition Control on HEX.

## Intended to use:

RT-PCR based diagnostic kit for the quantitative detection of Chicken from a high level of food and feed safety, accurate animal species identification and the detection of adulterants are two of the greatest challenges facing food and feed products companies today.. It is recommended to use standard Tissue DNA extraction kit (spin column based or magnetic bead based) for isolation of Chicken DNA. The kit will also work on the automated DNA purification system. The kit intended to use by trained laboratory technician.

## Contents of Kit:

| Components               | Color Code | Volume |
|--------------------------|------------|--------|
| RT Master Mix (MM)       | Natural    | 1000µL |
| PP Mix                   | Amber Tube | 200µL  |
| Positive control (PC)    | Blue       | 100µL  |
| Nuclease free water (NC) | White      | 100µL  |

All Vials have Color Coder tops to distinguish between different reagents.

## Storage:

The kit is shipped and stored at -200C

## ❖ Sample which can be used for extraction:

Chicken (Tissue)

## Procedure:

The performance of the SN Chicken RT-PCR Detection Kit depends on the amount and quality of template DNA purified from the tissue samples. The extraction protocol should be performed following manufacturer's instructions or an internally validated protocol.

## SN PCR Master Mix preparation:

Prepare the master mix as per table mentioned below. Positive control (PC) and Negative test control (NC/NTC) should be included in every set up.

| Color Code   | Components         | Volume (μL)/Reaction | Volume (μL) for 100 Reaction |
|--------------|--------------------|----------------------|------------------------------|
| Natural      | SN Master Mix (MM) | 10 μL                | 1000 μL                      |
| Amber Tube   | PP Mix             | 2 μL                 | 200 μL                       |
| Total Volume |                    | 12 μL                | 1200 μL                      |

Add each component to multiple numbers of samples to be processed and prepare the master mix. It is advisable to prepare extra mix for one sample for any pipetting related error or dead volumes.

## Note:

- ❖ Dispense 12 μL from above master mix in separate tubes/wells
- ❖ Add 4 μL of isolated DNA from sample and 4 μL of nuclease free water separately to each tube and total reaction mixture volume is 20 μL

Once dispensed, seal the plate with optically clear sealers or close the tubes-strips, centrifuge the tubes/strips or plates briefly and place them in the instrument.



## Thermal Profile: (Setting of RT-PCR)

Referring to the instrument manual, set on the dedicated software for the parameters of thermal cycle.

| Steps   | Temp  | Time    | Cycle |
|---------|-------|---------|-------|
| Hold 1  | 95 °C | 20 sec  | 1     |
| Cycling | 95 °C | 1 min   | 40    |
|         | 60 °C | 20 secs |       |
|         | 72°C  | 05 sec  |       |

## Fluorescence Setup:

| Applied Biosystem 7500 | BioRad CFX-96 | Rotor Gene Q  |
|------------------------|---------------|---------------|
| Cy5                    | Cy5           | Red Channel   |
| IC                     | HEX           | Green Channel |

### Note:

The Kit is compatible with; Rotor Gene™ 3000/6000, ABI Step One, 7500/FAST, Quant studio, BioRad CFX 96, Roche 480 and other Real Time PCR machine available.

## Precautions for PCR:

**The following aspects should always be taken care of:**

- Store positive material (Positive controls & amplicons) separately from all other reagents and add it to the reaction mix in a separate area.
- Thaw all components thoroughly at room temperature before starting the assay.
- When thawed, mix the components and centrifuge briefly.
- Work quickly on ice or in the Cooling Block.
- All the reagents including the NTC (except for PCs & specimens) should be mixed & dispensed in pre-mix area.
- All the controls & specimens should be mixed & dispensed in extraction area.
- Use pipette tips with filters only.

- h) Always use disposable powder-free gloves.
- i) Discard samples and assay waste according to your local safety regulations.
- j) Do not open the reaction tubes or plates post amplification.

### Additionally, required Materials and Devices

- a) Appropriate 3 or more channels RT-PCR instrument, compatible with the fluorophores used in this test.
- b) Appropriate nucleic acid extraction system or kit. It is recommended to use commercialized extraction kit extracting DNA.
- c) Vortex mixer.
- d) Centrifuge with a rotor for 2 mL micro centrifuge tubes.
- e) Pipette tips with filters (disposable).
- f) Powder-free gloves (disposable).
- g) Class II Biological Safety Cabinet (BSC), ideally in a BSL-2 facility.

### Result Interpretation:

Amplification of PC suggests absence of any inhibition during amplification. Amplification of IC in samples suggests proper DNA extraction. Amplification in Cy5 channels indicates presence of Chicken in sample. Due to competition and limiting factors, samples with high copy may not show amplification in HEX (IC) channel and the curves may not be a proper sigmoid. Results can be analyzed with the help as mentioned below:

#### Note:

Ct value in any sample /well >36 should be considered as below detection limit of the kit

### Result Interpretation table:

| S.NO | HEX<br>(internal control) | Cy5 | Result Interpretation           |
|------|---------------------------|-----|---------------------------------|
| 1.   | +                         | +   | Chicken tissue DNA detected     |
| 2.   | +/-                       | -   | Chicken tissue DNA Not detected |

|    |     |   |                                                    |
|----|-----|---|----------------------------------------------------|
| 3. | +/- | + | Presumptive Chicken tissue DNA detected*           |
| 4. | +   | - | Targets are valid, Chicken tissue DNA NOT detected |
| 5  | -   | - | Invalid repeat sampling/extraction/amplification   |

\*Retest the sample if the retest results are same, then it could be presumed as positive.

## Analytical Performance:

Evaluation of the kit on the confirm positives and negative specimen showed 100% specificity and sensitivity. The kit can detect  $1 \times 10^3$  copies per ml of Chicken DNA from tissue sample.

## Trouble shooting:

| Problem Possible Cause Recommendation               |                                                                                                                         |  |                                                                                                                                                                                                                                                                                                         |
|-----------------------------------------------------|-------------------------------------------------------------------------------------------------------------------------|--|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
|                                                     | Error in Master mix preparation.                                                                                        |  | Check the dispensing volume during preparation of master mixture                                                                                                                                                                                                                                        |
| No signal in all samples including positive control | Inhibitors added<br>Probe degradation<br>Positive control degradation<br>Omitted components<br>Instrument setting error |  | Repeat the extraction step with new sample<br>Use a new probe reagent<br>Use a new positive control<br>Verify each component, repeat the RT-PCR mixture preparation<br>Check the position setting for the positive control on the instruments.<br>Check the Thermal cycle settings on sample instrument |
| Diverse intensity of fluorescent signals            | Pipetting error<br>Contamination in the outer surface of PCR tubes or Plate                                             |  | Make sure that the equal volume of reactants is added in each tube or plate<br>Wear gloves during the experiment                                                                                                                                                                                        |
| Weak or no fluorescent signal in samples only       | Poor DNA quality<br>Insufficient volume of DNA                                                                          |  | Use recommended kit for DNA extraction and store extracted DNA at -70°C<br>Repeat PCR reaction with correct                                                                                                                                                                                             |



|                                                       |                                                                                                                                                                                                        |                                                                                                                                 |
|-------------------------------------------------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|---------------------------------------------------------------------------------------------------------------------------------|
| <b>No template Control(NTC) showing Amplification</b> | Amplification of Chicken genes in a no Template Control indicates contamination in one or more of the reagents, incorrect placement of a plate or tubes into the RT-PCR instrument, or pipetting error | Repeat the test by using 2 NTC. If NTC fails again, then investigation should be conducted to identify possible causes of error |
|-------------------------------------------------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|---------------------------------------------------------------------------------------------------------------------------------|

### **Technical Assistance:**

For technical assistance please contact our technical Support:

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